

ARRAY 4



ARRAY 4 – Antibody

GLUTEN-ASSOCIATED
CROSS-REACTIVE FOODS
& FOODS SENSITIVITY™

TABLE OF CONTENTS

Overview

Influencing Factors

Genetic

Environmental

History

Clinical – Systemic Immune Effects

Clinical Use of Antibody Array 4

Clinical Interpretation of Antibody Array 4

Specimen Requirement

Related Testing

References

GLUTEN-ASSOCIATED CROSS-REACTIVE FOODS AND FOODS SENSITIVITY™

OVERVIEW

Once a patient is diagnosed as Celiac Disease (CD) or having Gluten Reactivity, he/she is instructed to adhere to a gluten-free diet. Brochures, books and websites help the patient with this seemingly difficult process. However, a significant percentage of these patients will continue to have gluten-like complaints even after being on a gluten-free diet (GFD) for months. Most countries define “gluten-free” products based on the recommendation of the Food and Agricultural Organization of the United Nations and World Health Organization. This codex alimentarius allows the inclusion of up to 0.3% protein from gluten containing grains in foods labeled “gluten-free.” If the sensitive body is exposed to 0.3% protein, the immune system will recognize and react to the protein.

There exists antigenic similarity, or cross-reaction, among many grains, and other dietary proteins such as casein with gluten. Based on biological individuality of immune response against a repertoire of gliadin or gluten peptides, any number of patients may produce antibodies against a single gluten antigen or a combination of gluten antigens, some of which may be cross-reactive with other food antigens.

A problem with digesting dairy, casein in particular,¹ may be a feature in about 50% of patients with CD and may, therefore, contribute to persistent symptoms in patients who are on a GFD.

Additionally, patients who are new to the GFD encounter new foods and/or over-consume old favorites to compensate for the lack of wheat in the diet. Gluten-free cookies, crackers, breads and cakes often contain copious amounts of rice, amaranth, sorghum and other substitutes. Some of these new-to-the-patient foods may illicit an adverse reaction. Other foods that are often introduced to the patient on the GFD are quinoa, buckwheat and hemp. Some patients may turn to the “ancient” grains (Polish wheat, spelt, barley, rye), not knowing that these contain gluten. Another problem patients often face on the GFD is the over-consumption of another starch to make up for the loss of wheat. They turn to potato, rice or corn as a substitute. This can lead to the development of a new sensitivity or the enhancement of old sensitivities.

Array 4 can assist the clinician by detecting both sensitivities and cross-reactions and thus reveal the possible cause of this continued gluten-like reaction in the patient.

Antibody Array 4

Testing for Gluten-Associated Cross-Reactive Foods and Foods Sensitivity in this array can assist the clinician in revealing the possible cause of this continued gluten-like reaction in the patient. Patients with Gluten-Reactivity or CD are sensitized to a broad range of dietary proteins, due to enzyme dysfunction, villi damage, or other disorders. Therefore, it is crucial to identify not only sensitivities to foods that are often recommended for patients on the GFD, but also the food antigens that cross-react to gluten peptides in the patient. Without biochemically individualized dietary intervention, the Gluten-Reactive patient may develop additional sensitivity/intolerance and autoimmunity.

[Top](#)

Although the majority of individuals with CD have substantial improvement within the first few weeks of gluten withdrawal, between 7% and 30% continue to have symptoms or clinical manifestations suggestive of CD despite being on a gluten-free diet.²

Non-responsive Celiac Disease (NRCD) is defined as:

- (1) Referral to a clinician specializing in CD for the evaluation of a lack of response to a gluten-free diet
- (2) Failure of clinical symptoms or laboratory abnormalities typical of CD to improve within 6 months of gluten withdrawal
- (3) Recurrence of symptoms and/or laboratory abnormalities typical of CD while on a gluten-free diet. Of the 12 identified causes of NRCD, the most common cause was (inadvertent) gluten exposure, accounting for 36% of patients.³ What about the other 64%? An all-too-common contributor to NRCD is cross-reactivity with other foods. Antibody cross-reactivity between different foods or between food and aeroallergens, such as trees and grasses, occurs much more readily than clinically evident cross-reactivity.⁴ The patient often is unable to ‘feel’ the immune response to cross-reactive food.

These are the confusing scenarios when a Gluten-Reactive person will say, “What did I eat that was a problem? The packaging didn’t reference any wheat products.” This can be explained in the following:

1. **Consumption of gluten-containing foods such as beer or chewing gum, and grains, such as spelt or barley** - Most countries define “gluten-free” products based on the recommendation of the Food and Agricultural organization of the United Nations and World Health Organization, which allows the inclusion of up to 0.3% protein from gluten-containing grains in foods labeled “gluten-free.”⁵ Wheat/gliadin is commonly added to foods as a “hidden” ingredient such as “Natural Flavor” or “Spices.” A complete understanding of ‘hidden wheat/gluten’ sources is necessary to have a better outcome with the GFD. Additionally, education of gluten-containing grains, such as rye,^{6 7} barley, ^{6 7} Polish wheat,^{7 8 9} and spelt.^{7 10 11 12 13}
2. **Problem with digesting dairy, in particular, casein sensitivity to cow’s, sheep’s and goat’s milk.** Casein sensitivity may be a feature in about 50% of patients with CD and may, therefore, contribute to persistent symptoms in Celiac patients who are on a gluten-free diet.^{1 14} Casein also has been suggested as an environmental trigger of other autoimmune disorders such as Behçet’s disease, type-1 diabetes, and systemic lupus erythematosus.^{15 16 17}
3. **Cross-reaction among foods, or infectious agents, and products with gluten.** For example, milk, casein, yeast and many other, as-yet-unidentified, food antigens, *salmonella typhi*, rotavirus and many other infectious agents, human tissue antigens, such as transglutaminase, heat shock protein, myotubularin-related

[Top](#)

protein 2 and cell surface receptors (toll-like receptors), all cross-react with gliadin or Celiac peptide.^{14 18 19 20} Indeed, bovine milk caseins and transglutaminase-treated cereal prolamins, such as wheat and maize, are differentially recognized by IgA of CD patients.²¹ Studies have identified food antigen cross-reactivity such as milk chocolate,^{22 23} rye,^{6 7} barley,^{6 7} Polish wheat,^{7 8 9} spelt,^{7 10 11 12 13} millet,^{23 64} corn,²³ rice,²³ yeast,^{23 24 25 26 27 28} oats,^{29 30 31 32 33 34 35} instant coffee.^{23 36 37 38 39 40} The response to some of these food allergens parallels the response to gliadin and might be relevant to the pathogenesis of Gluten-Reactivity and CD by increased mucosal permeability, resulting in elevated antigen reabsorption, which can trigger immune activation with elevated levels of IgA antibodies.⁴¹ Perhaps this is why as many as 40% of children on a well-managed gluten-free diet for at least 1 year still have elevated antibodies to gluten.⁴²

- 4. Consumption of new foods and over-consumption of alternative starches.** Patients new to the GFD often begin to consume foods never eaten as a child when humans develop tolerance to foods. As an adult on a GFD, some of these gluten-alternatives (sesame,^{43 44 45 46 47} hemp,⁴⁸ buckwheat,^{49 50 51 52 53 54 55 56 57} sorghum,^{58 59 60 61 62} millet,^{63 64 65 66} amaranth,^{67 68 69} quinoa,^{70 71 72} tapioca,^{73 74 75 76} teff^{77 78}) may cause the immune system to react against the new food as if it were a foreign antigen. Another common event is for a patient to develop food-sensitivity due to over-consumption of a specific food. People new to the GFD may substitute another starch to compensate for the loss of wheat in the diet. Thus, the development of reactions to corn,^{79 80 81} rice,^{82 83 84 85 86} and/or potato^{87 88 89} is seen in some patients on a GFD.
- 5. Common food sensitivities.** The gluten-reactive patient may also be reactive to common food antigens, such as hen's eggs^{90 91 92} and soybean.^{93 94}

From these data, it is conceivable that patients with Celiac disease are sensitized to a broad range of dietary proteins and peptides. Therefore, it is crucial to identify food antigens with a capacity to sensitize patients with Celiac and other autoimmune disorders.^{1 5 95 96}

Negative serology for transglutaminase, endomysium, or gliadin should not necessarily reassure the clinician⁹⁷ of neither negative immune activation nor pathology from Gluten Reactivity. Several reports^{98 99 100 101 102} show that in the majority of Celiac patients, these antibodies may be negative or low but cross-reactive antibodies could be elevated. From the diagnostic and therapeutic point of view, it makes sense to define allergen clusters (cross-reactivity).¹⁰³

INFLUENCING FACTORS

GENETIC

Close to 90% of CD patients carry the gene DQ2 (*DQA1*05/DQB1*02*), and a minority (10%) of the CD patients carry DQ8 (*DQA1*03/DQB1*0302*). Typically, gluten peptides bind to the DQ2 and DQ8 molecules. Recent research however, has identified at least eight new genomic regions with robust levels of disease association to Gluten-Reactivity.^{104 105}

[Top](#)

ENVIRONMENTAL (CHEMICALS, FOODS, BIOTOXINS, DRUGS...)

Environmental factors that have an important role in the development of CD have been suggested by epidemiologic studies. These include a protective effect of breast-feeding¹⁰⁶ and the introduction of gluten in relation to weaning.^{107 108}

Numerous environmental factors have been hypothesized as being catalysts for the development of not only the gluten enteropathy CD,¹⁰⁹ but also systemic manifestations of Gluten-Reactivity with or without the enteropathy. Some of these catalysts include bacteria,¹¹⁰ viruses,¹⁸ gut dysbiosis,¹¹¹ and cross-reactive foods.¹¹²

HISTORY (FAMILY, MEDICAL)

CD and Gluten-Reactivity are characterized by a variety of clinical manifestations. These include the typical malabsorption syndrome (classic symptoms) and a spectrum of symptoms potentially affecting any organ or body system (non-classic symptoms).^{113 114 115}

Clinical manifestations of Gluten-Reactivity and CD along with sensitivity to cross-reactive foods can present at any age:

- **Infancy** (less than 2 years old) – diarrhea, abdominal distention, failure to thrive (low weight, lack of fat, hair thinning), anorexia, vomiting, psychomotor impairment (muscle wasting)
- **Childhood** – diarrhea, constipation, anemia, loss of appetite, short stature, osteoporosis
- **Adulthood** – diarrhea, constipation, anemia, aphthous ulcers, sore tongue & mouth (mouth ulcers, glossitis, stomatitis), dyspepsia, abdominal pain, bloating (weight loss), fatigue, infertility, neuropsychiatric symptoms (anxiety, depression, etc.), bone pain (osteoporosis), weakness (myopathy, neuropathy).^{116 117 118}

Reviewing current medications (antibiotics, steroids, NSAID's, etc.), supplements, diets, and a detailed medical history are critically important in determining who may have gluten sensitivity. The correlation between food ingestion and symptom onset is of great clinical importance.

CLINICAL – SYSTEMIC IMMUNE EFFECTS

When cross-reactivity is present in a patient, gluten antibodies may be essentially normal, and antibodies to the particular antigenic food may be the sole indicator of a continued inflammatory response, triggering the symptomatology of CD.

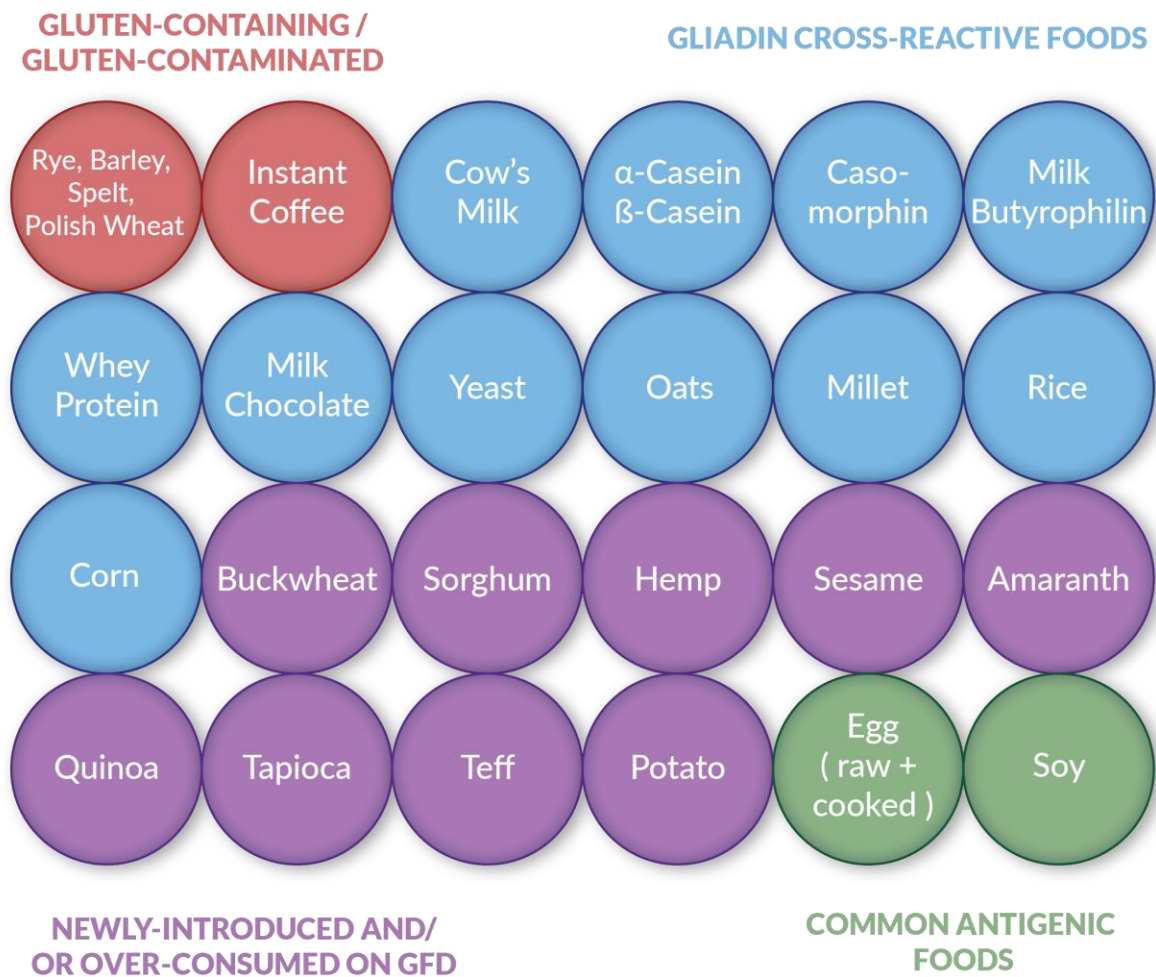
Reduced antibodies to gluten—after introduction of a gluten-free and cross-reactive food diet—probably reflects catabolism of pre-formed antibodies combined with lowered synthesis due to the lack of antigen stimulation. Concurrent reduction of antibodies to other dietary antigens may, therefore, be a better indication of improved mucosal integrity by reflecting decreased penetrability of antigens still available in the gut lumen.

Determination of serum IgA and IgG antibody activities to dietary proteins appears to be a valuable adjunct in the diagnosis and follow-up of diagnosed CD, both in children and adults. Increased IgA activities to other dietary antigens are likewise relatively characteristic of untreated

CD; monitoring of such antibodies may be particularly helpful in evaluating the response of patients on a gluten- and cross-reactive food-free diet.¹¹⁹

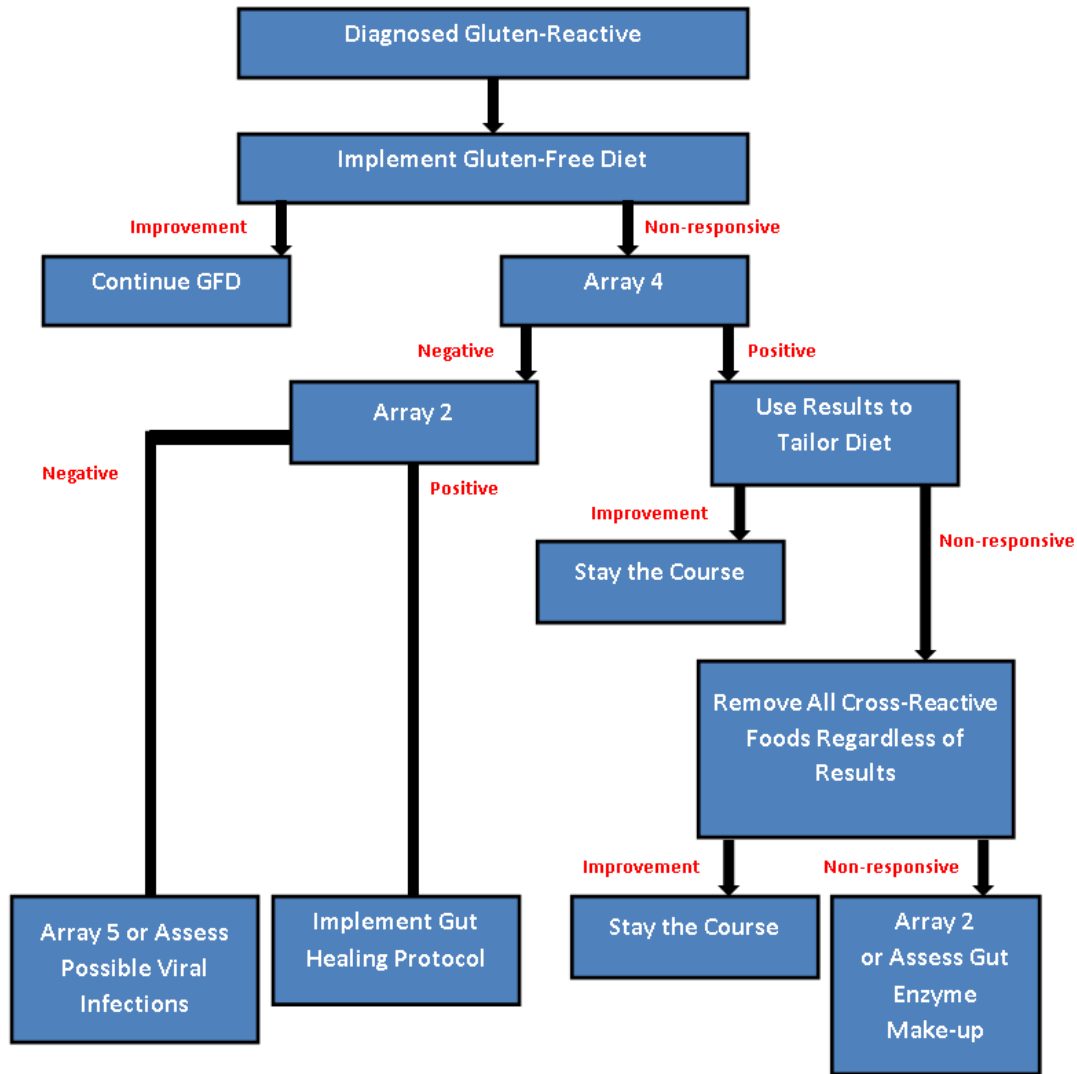
The manifestations and the pathophysiology of CD and Gluten-Reactivity can be as unique as the individual himself. Identifying these triggers and cascades of autoimmunity is an important step in designing effective treatment and maintenance protocols for the patient. Therefore, patients with CD or Gluten-Reactivity suffer an array of autoimmunity beyond the gastrointestinal system.

Cyrex Laboratories' Antibody Arrays for Gluten-Reactivity are vital components to clinical practice. After establishing the patient on a gluten-free diet, many will return after adhering to this diet for months, and yet they still exhibit the same clinical complaints as they experienced with gluten-containing foods. Undoubtedly, these patients are having reactions to foods which cross-react with gluten antigens. Antibody Array 4 – Gluten-Associated Cross-Reactive Foods and Foods Sensitivity is designed to assess these select individuals. With results of this array, the practitioner can take a better, broader approach to developing a tailored diet plan for patients with Celiac disease or gluten sensitivity.



[Top](#)

CLINICAL USE OF ANTIBODY ARRAY 4



Patients with Gluten-Reactivity and CD are sensitized to a broad range of dietary proteins due to enzyme dysfunction, villi damage, or other disorders. A common problem is the digestion of dairy products, the casein protein, in particular. Consuming these food products will cause persistent symptoms and clinical complaints similar to the initial discomforts of the gluten sensitivity.

Complete normalization of gut lesions is very rare in adult patients with Celiac disease (8%), despite gluten-free diet compliance. Although a majority (65%) feels better, the ensuing inflammation in the gastrointestinal tract, due to cross-reactions with – and sensitization to – an array of food antigens, remains a cause for clinical concern. When the patient, despite adamant adherence to the gluten-free diet, is non-responsive, continues to exhibit clinical complaints or has therapy-resistant gut dysbiosis, an assessment of IgG + IgA antibodies to an array of food

antigens associated with a gluten-free diet, or known to cross-react with gluten, can guide the Healthcare Practitioner in tailoring a recovery diet plan and preventing devastating autoimmune disorders.

Gluten-Associated Cross-Reactive Foods and Foods Sensitivity assessment is recommended for patients who:

- Have Gluten-Reactivity or Celiac disease
- Are non-responsive to the gluten-free diet
- Have gut dysbiosis, which appears to be resistant to standard therapy
- Have an autoimmune disorder

[Top](#)

CLINICAL INTERPRETATION OF ANTIBODY ARRAY 4

Foods Known to Cross-React With Purified Alpha-Gliadin-33-Mer	
Cow's Milk	Gluten Grains*
$\alpha + \beta$ Casein	Yeast
Casomorphin	Oats
Milk Butyrophilin	Millet
Whey Protein	Rice
Milk Chocolate	Corn

* Rye, Barley, Spelt, Polish Wheat (Polish Wheat is also known as Camel's wheat, Egyptian wheat, Khorasan wheat and Kamut®)

For patients with known gluten reactions or Celiac disease (refer to results from Array 1 or Array 3), all cross-reactive foods should be removed from the patient's diet under a clinician's care.

Newly-Introduced and Over-Consumed Foods on GFD	
Sesame	Quinoa
Buckwheat	Tapioca
Sorghum	Teff
Millet*	Rice*
Hemp	Corn*
Amaranth	Potato

**Also cross-reacts with alpha-gliadin-33-mer*

If any foods commonly consumed on the gluten-free diet result positive, the offending food should be eliminated from the patient’s diet until the gut is healed. Slowly reintroduce the foods on a rotation diet after gut is healed. After at least two months on the rotation diet, Array 4 may be rerun on a fresh specimen.

[Top](#)

INTERPRETATION OF CROSS-REACTIVE AND FOOD ANTIGENS						
POSITIVE REACTION TO:	DAIRY REACTIVITY	IN VITRO CROSS-REACTION TO α -GLIADIN	NEWLY INTRODUCED FOODS ON GFD	OVER-CONSUMED ON GFD	GLUTEN-CONTAINING/ GLUTEN-CONTAMINATED	COMMON ANTIGENIC FOODS
Rye, Barley, Spelt, Polish Wheat*						
Instant Coffee						
Cow's Milk						
α -Casein + β -Casein						
Casomorphin						
Milk Butyrophilin						
Whey Protein						
Milk Chocolate						
Yeast						
Oats						
Millet						
Rice						
Corn						
Sesame						
Buckwheat						
Sorghum						
Hemp						
Amaranth						
Quinoa						
Tapioca						
Teff						
Potato						
Soy						
Egg						

*Polish Wheat is also known as Camel's wheat, Egyptian wheat, Khorasan wheat and Kamut®

Dairy Reactivity – if any of the 6 antigens are positive, the patient must be placed on a dairy-free diet.

Gluten-Containing Grains – if the patient is wheat-sensitive, but not gluten-sensitive, he/she may be able to tolerate these grains. If any of these are positive the patient must be on a gluten-free diet. If the patient is already on a long-term (more than four months) GFD and is positive for gluten-containing grains, he/she is consuming 'hidden' sources of gluten.

In Vitro Cross-Reaction to Gliadin – in the laboratory setting these foods were shown to be highly cross-reactive to purified gliadin. If the patient had antibodies to any of these foods based on the practitioner's recommendation, the foods should be eliminated from the patient's diet.

Newly Introduced Foods – when a patient goes on a gluten-free diet there are many exposures of foods the patient may not have eaten during the formative years, when humans develop their tolerance to foods, thus, the patient may have an adverse reaction to the new food. Positive antibodies to these foods means the foods should be avoided. After normalization of the immune

response, these foods may be reintroduced on a rotation basis and rechecked for reactivated immune response after a minimum of 2 months fresh exposure.

Over-Consumed Foods – when a patient goes on a gluten-free diet, the patient often trades one sensitivity for another by over eating a different starch. The common substitutes tend to be potato, rice or corn. After normalization of the immune response, these foods may be reintroduced on a rotation basis and rechecked for reactivated immune response after a minimum of 2 months fresh exposure.

Wheat-Contaminated Foods – although oats in a recent study did not cross-react with purified alpha-gliadin-33-mer,²³ researchers found that certain varieties of oats do cross-react with gliadin.^{31 35} It is recommended that gluten reactive patient's refrain from consuming oats not only because one does not know the variety of oats in the package purchased at the grocery store, but also due to the common wheat contamination of oats that occurs in the transportation, processing and packaging processes.³³ The coffee antigen used on Array 4 is instant coffee. In a recent study,²³ instant coffee was shown to cross-react with gliadin, but whole bean coffee was not. This indicates that coffee protein by itself does not cross-react with purified gliadin. The positive result for instant coffee shows that during the manufacturing of instant coffee, wheat is either added to the product or contaminates it. Gluten-reactive patient who test positive to instant coffee on Array 4 should eliminate all coffee for a period of time to heal the gut and quiet the immune response. Whole bean coffee can be reintroduced and the patient monitored for adverse reactions.

TREATMENT PROTOCOL

1. Tailor a more effective, individualized diet plan. Refer to antigen specification sheets for additional known cross-reactions for the best dietary outcome. Specification sheets are found under “Tests and Arrays” then click “Array 4” followed by the pdf symbol next to the food antigen.
2. Heal the gut.
3. After confirmation that the gut is healed, using a rotation diet, slowly re-introduce the non-cross-reactive foods into the patient's diet regimen.
4. After fully re-introducing the foods, retest with Array 4.
5. If antibody levels have normalized, continue with the rotation plan.
6. If antibody levels have not normalized, instruct the patient to avoid the positive foods for life.

Specimen Requirement

2 mL Serum

Ambient

Related Testing

Antibody Array 2 – Intestinal Antigenic Permeability Screen (Serum)

[Top](#)

REFERENCES

- ¹ Kristjánsson G, Venge P, Hällgren R. Mucosal reactivity to cow's milk protein in Coeliac disease. *Clin Exp Immunol*. 2007; 147(3):449-455.
- ² Green P and Cellier C. Celiac Disease. *N Engl J Med*, 2007; 357:1731-1743.
- ³ Leffler DA, Dennis M, Hyett B, *et al.* Etiologies and predictors of diagnosis in nonresponsive Celiac disease. *Clin Gastroenterol Hepatol*, 2007; 5(4):445-450.
- ⁴ Eckman J, Saini SS, Hamilton RG. Diagnostic evaluation of food-related allergic diseases. *Allergy Asthma Clin Immunol*, 2009; 5(1):2.
- ⁵ Faulkner-Hogg KB, Selby WS, Loblay RH. Dietary analysis in symptomatic patients with Coeliac disease on a gluten-free diet: the role of trace amounts of gluten and non-gluten food intolerances. *Scand J Gastroenterol*, 1999; 34:384-389.
- ⁶ Ciclitiera PJ and Ellis HJ. Relation of antigenic structure of cereal proteins to their toxicity in Coeliac patients. *Brit J Nutr*, 1985; 53:39-45.
- ⁷ Weber D, Cléroux C, Benrejeb Godefroy S. Emerging analytical methods to determine gluten markers in processed foods – method development in support of standard setting. *Anal Bioanal Chem*, 2009; 395:111-117.
- ⁸ Kasarda DD. Grains in relation to Celiac disease. *Cereal Foods World*, 2001; 46:209-210.
- ⁹ Simonato B, Pasini G, Giannattasio M, Curioni A. Allergenic potential of Kamut® wheat. *Allergy*, 2002; 57:653-654.
- ¹⁰ Grela ER. Nutrient composition and content of antinutritional factors in spelt (*Triticum spelta* L.) cultivars. *J Sci Food Agric*, 1996; 71(3):399-404.
- ¹¹ Jones SM, Megnolfi CG, Cooke SK, Sampson HA. Allergens, IgE, mediators, inflammatory mechanisms: immunologic cross-reactivity among cereal grains and grasses in children with food hypersensitivity. *J Allergy Clin Immunol*, 1995; 96:341-351.
- ¹² Pastorello EA, Farioli L, Robino A, *et al.* A lipid transfer protein involved in occupational sensitization to spelt. *J Allergy Clin Immunol*, 2001; 108(1):145-146.
- ¹³ Skrabagnja V, Kovac B, Golob T, *et al.* Effect of spelt wheat flour and kernel on bread composition and nutritional characteristics. *J Agric Food Chem*, 2001; 49:497-500.
- ¹⁴ Wildner G and Diedrichs-Möhrling M. Autoimmune uveitis induced by molecular mimicry of peptides from rotavirus, bovine casein and retinal S-antigen. *Eur J Immunol*, 2003; 33:2577-2587.
- ¹⁵ Triolo G, Accardo-Palumbo A, Dieli F, *et al.* Humoral and cell-mediated immune response to cow's milk proteins in Behcet's disease. *Ann Rheum Dis*, 2002; 61:459-462.

[Top](#)

- ¹⁶ Monetini L, Barone F, Stefanini L, *et al.* Establishment of T-cell lines to bovine beta-casein and beta-casein-derived epitopes in patients with type 1 diabetes. *J Endocrinol*, 2003; 176:143-150.
- ¹⁷ Riemekasten G, Marell J, Hentschel C, *et al.* Casein is an essential cofactor in autoantibody reactivity directed against the C-terminal SmD1 peptide AA83-119 in systemic lupus erythematosus. *Immunobiology*, 2002; 206:537-545.
- ¹⁸ Zanoni G, Navone R, Lunardi C, *et al.* In Celiac disease, a subset of autoantibodies against transglutaminase binds to toll-like receptor 4 and induces activation of monocytes. *PLoS Med*, 2006; 3(9):1637-1653.
- ¹⁹ Blutt SE, Crawford SE, Warfield KL, *et al.* The VP7 outer capsid protein of rotavirus induces polyclonal B-cell activation. *J Virol*, 2004; 78:6971-6981.
- ²⁰ Sollid LM and Gray GM. A role for bacteria in Celiac disease? *Am J Gastroenterol*, 2004; 99:905-906.
- ²¹ Cabrera-Chávez F, Rouzaud-Sáñez O, Sotelo-Cruz N, *et al.* Bovine milk caseins and transglutaminase-treated cereal prolamins are differentially recognized by IgA of Celiac disease proteins according to their age. *J Agric Food Chem*, 2009; 57:3754-3759.
- ²² Becker CG, Van Hamont N, Wagner M. Tobacco, cocoa, coffee, and ragweed: cross-reacting allergens that activate factor-XII-dependent pathways. *Blood*, 1981; 58(5):861-867.
- ²³ Vojdani A and Tarash I. Cross-reaction between gliadin and different food and tissue antigens, *Food Nutri Sci*, 2013; 4:20-32.
- ²⁴ Heelan BT, Allan S, Barnes RMR. Identification of a 200-kDa glycoprotein antigen of *Saccharomyces cerevisiae*. *Immunol Lett*, 1991; 28:181-186.
- ²⁵ Oshitani N, Hato F, Kenishi S, *et al.* Cross-reactivity of yeast antigens in human colon and peripheral leukocytes. *J Pathol*, 2003; 199:361-367.
- ²⁶ Sendid B, Quinton JF, Charrier G, *et al.* Anti-Saccharomyciescerevisiaemannan antibodies in familial Crohn's disease. *Am J Gastroenterol*, 2001; 93(8):1306-1310.
- ²⁷ Vojdani A, Rahimian P, Kalhor H, Mordechai E. Immunological cross reactivity between candida albicans and human tissue. *J Clin Lab Immunol*, 1996; 48:1-15.
- ²⁸ Young CA, Sonnenberg A, Berns, EA. Lymphocyte proliferation response to baker's yeast in Crohn's disease. *Digestion*, 1994; 55(1):40-43.
- ²⁹ Arentz-Hansen H, Fleckenstein B, Molberg Ø, *et al.* The molecular basis for oat intolerance in patients with Celiac disease. *PLoS Med*, 2004; 1(1):84-92.
- ³⁰ Janatuinen EK, Pekka HP, Kempainen TA, *et al.* A comparison of diets with and without oats in adults with Celiac disease. *N Engl J Med*, 1995; 333:1033-1037.

[Top](#)

- ³¹ Reunala T, Collin P, Holm K, *et al.* Tolerance to oats in dermatitis herpetiformis. *Gut*, 1998; 43:490-493.
- ³² Silano M, Dessì M, De Vincenzi M, Cornell H. In Vitro tests indicate that certain varieties of oats may be harmful to patients with Coeliac disease. *J Gastroenterol Hepatol*, 2007; 22:528-531.
- ³³ Srinivasan U, Jones E, Carolan J, Feighery C. Immunohistochemical analysis of Coeliac mucosa following ingestion of oats. *Clin Exp Immunol*, 2006; 144:197-203.
- ³⁴ Thompson T. Gluten contamination of commercial oat products in the United States. *N Engl J Med*, 2004; 351(19):2021-2022.
- ³⁵ Comino I, Real A, de Lorenzo L, *et al.* Diversity in oat potential immunogenicity: basis for the selection of oat varieties with no toxicity in coeliac disease. *Gut*, 2011;
- ³⁶ Axelsson IG. Allergy to the coffee plant. *Allergy*, 1994; 49(10):885-887.
- ³⁷ Caballero T, Garcia-Ara C, Pascual C, *et al.* Urticaria induced by caffeine. *J Investig Allergol Clin Immunol*, 1993; 3(3):160-162.
- ³⁸ Moneret-Vautrin DA, Kanny G, Faller JP, *et al.* Severe anaphylactic shock with heart arrest caused by coffee and gum Arabic, potentiated by beta-blocking eyedrops. (Article in French) *Rev Med Interne*, 1993; 14(2):107-111.
- ³⁹ Osterman K, Johansson SG, Zetterström O. Diagnostic tests in allergy to green coffee. *Allergy*, 1995; 40(5):336-343.
- ⁴⁰ Treudler R, Tebbe B, Orfanos CE. Coexistence of type I and type IV sensitization in occupational coffee allergy. *Contact Dermatitis*, 1997; 36:109.
- ⁴¹ Hvatum M, Scott H, Brandtzaeg P. Serum IgG subclass antibodies to a variety of food antigens in patients with Coeliac disease. *Gut*, 1992; 33(5):632-638.
- ⁴² Husby S, Foged N, Oxelius VA, Svehag SE. Serum IgG subclass antibodies to gliadin and other dietary antigens in children with Coeliac disease. *Clin Exp Immunol*, 1986; 64(3):526-35.
- ⁴³ Gangur V, Kelly C, Navuluri L. Sesame allergy: a growing food allergy of global proportions? *Ann Allergy Asthma Immunol*, 2005; 95:4-11.
- ⁴⁴ Kagi MK and Wuthrich B. Falafel burger anaphylaxis due to sesame seed allergy. *Ann Allergy*, 1993; 71(2):127-129.
- ⁴⁵ Keskinen H, Ostman P, Vaheria E, *et al.* A case of occupational asthma, rhinitis and urticaria due to sesame seed. *Clin Exp Allergy*, 1991; 21:623-624.
- ⁴⁶ Pecquet C, Leynadier F, Saïag P. Immediate hypersensitivity to sesame in foods and cosmetics. *Contact Dermatitis*, 1998; 39:313.
- ⁴⁷ Perkins MS. Raising awareness of sesame allergy. *Pharma J*, 2001; 267:757-758. [Top](#)

- ⁴⁸ Popa V, Gavrilesco N, Preda N, *et al.* An investigation of allergy in byssinosis: sensitization to cotton, hemp, flax and jute antigens. *Brit J Industr Med*, 1969; 26:101-108.
- ⁴⁹ Göhte C-J, Wislander G, Ancker K, Forsbeck M. Bucksheat allergy: health food, an inhalation health risk. *Allergy*, 2007; 38(3):155-159.
- ⁵⁰ Hekkens WT. The determination of prolamins in gluten-free food. Introductory remarks. *Panminerva Med*, 1991; 33(2):61-64.
- ⁵¹ Kim J-L, Wieslander G, Norbäck D. Allergy/Intolerance to buckwheat and other food products among Swedish subjects with Celiac disease. *Proc. 9th Int'l Symp Buckwheat*, Prague, 2004; 74:705-709.
- ⁵² Lee SY, Lee KS, Hong CH, Lee KY. Three cases of childhood nocturnal asthma due to buckwheat allergy. *Allergy*, 2001; 56:763-766.
- ⁵³ Pomeranz Y, Marshall HG, Robbins GS, Gilbertson JT. Protein content and amino acid composition of maturing buckwheat (*Fagopyrum esculentum* Moench). *Cereal Chem*, 1975; 52:479-484.
- ⁵⁴ De Maat-Bleeker F and Stapel SO. Cross-reactivity between bucksheat and latex. *Allergy*, 1998; 53:538-539.
- ⁵⁵ Sdepanian VL, Scaletsky ICA, Fagundes-Neto U, de Moraes MB. Assessment of gliadin in supposedly gluten-free foods prepared and purchased by Celiac patients. *J Ped Gastroenterol Nutr*, 2001; 32:65-70.
- ⁵⁶ Skerritt JH, Devery JM, Hill AS. Chemistry, Coeliac-toxicity and detection of gluten and related prolamins in foods. *Panminerva Med*, 1991; 33(2):65-74.
- ⁵⁷ Wieslander G and Norbäck D. Buckwheat allergy. *Allergy*, 2001; 56:703-704.
- ⁵⁸ Bietz JA. Cereal prolamins evolution and homology revealed by sequence analysis. *Biochem Genetics*, 1982; 20(11/12):1039-1053.
- ⁵⁹ Cicek M and Esen A. Structure and expression of a dhurrinase (β -glucosidase) from sorghum. *Plant Physiol*, 1998; 116:1469-1478.
- ⁶⁰ Mazhar H, Chandrashekar A, Shetty HS. Isolation and immunochemical characterization of the alcohol-extractable proteins (kafirins) of *Sorghum bicolor* (L.) Moench. *J Cereal Sci*, 1993; 17(1):83-93.
- ⁶¹ Taylor JRN, Schüssler L, van der Walt WH. Fractionation of proteins from low-tannin sorghum grain. *J Agric Food Chem*, 1984; 32:149-154.
- ⁶² Gaitan E, Cooksey RC, Legan J, Lindsay RH. Antithyroid effects *in vivo* and *in vitro* of vitexin: a C-glycosylflavone in millet. *J Clin Endocrinol Metab*, 1995; 80(4):114-1147.

[Top](#)

⁶³ Monteiro PV, Virupaksha,TK, Rao DR. Proteins of Italian millet: amino acid composition, solubility fractionation and electrophoresis of protein fractions. *J Sci Food Agric*, 1982; 33(11):1072-1079.

⁶⁴ Monteiro PV, Sudharhsna L, Ramachandra G. Japanese barnyard millet (*Echinochloafrumentacea*): protein content, quality and SDS-PAGE of protein fractions. *J Sci Food Agric*, 1988; 43(1):17-25.

⁶⁵ Parameswaran KP and Thayumanavan B. Homologies between prolamins of different minor millets. *Plant Foods Human Nutr*, 1995; 48:119-126.

⁶⁶ Parameswaran KP and Thayumanavan B. Isolation and characterization of a 20 kDprolamin from kodo millet (*Paspalumscrobiculatum*) (L.): homology with other millets and cereals. *Plant Foods Human Nutr*, 1997; 50:359-373.

⁶⁷ Aphalo P, Castellani, O.F., Martinez, E.N., Anón, M.C. "Surface physicochemical properties of globulin-P amaranth protein." *J Agric Food Chem*, 2004; 52:616-622.

⁶⁸ Gorinstein S, Delgado-Licon E, Pawelzik E, *et al.* Characterization of soluble amaranth and soybean proteins based on fluorescence, hydrophobicity, electrophoresis, amino acid analysis, circular dichroism, and differential scanning calorimetry measurements. *J Agric Food Chem*, 2001; 49:5595-5601.

⁶⁹ Vasco-Méndez NL and Paredes-López, O. Antigenic homology between amaranth glutelins and other storage proteins. *J Food Biochem*, 1995; 18(4):227-238.

⁷⁰ Aluko RE and Monu E. Functional and bioactive properties of quinoa seed protein hydrolysates. *J Food Sci*, 2003; 68(4):1254-1258.

⁷¹ Lee AR, Ng DL, Dave E, *et al.* The effect of substituting alternative grains in the diet on the nutritional profile of the gluten-free diet. *J Hum Nutr Diet*, 2009; 22:359-363.

⁷² Wright KH, Huber KC, Fairbanks DJ, Huber CS. Isolation and characterization of *Atriplexhortensis* and sweet *Chenopodium quinoa* starches. *Cereal Chem*, 2002; 79(5):715-719.

⁷³ Beezhold DH, Sussman GL, Liss GM, Chang NS. Latex allergy can induce clinical reactions to specific foods. *Clin Exp Allergy*, 1996; 26(4):416-422.

⁷⁴ Brehler R, Theissen U, Hohr C, Luger T. Latex-fruit syndrome: frequency of cross-reacting IgE antibodies. *Allergy*, 1997; 52:404-410.

⁷⁵ Ibero M, Castillo MJ, Pineda F. Allergy to cassava: a new allergenic food with cross-reactivity to latex. *J Investig Allergol Clin Immunol*, 2007; 17(6):409-412.

⁷⁶ Mikkola JH, Alenius H, Kalkkinen N, *et al.* Hevein-like protein domains as a possible cause for allergen cross-reactivity between latex and banana. *J Allergy Clin Immunol*, 1998; 102:1005-1012.

[Top](#)

- ⁷⁷ Bergamo P, Maurano F, Mazzarella G, *et al.* Immunological evaluation of the alcohol-soluble protein fraction from gluten-free grains in relation to celiac disease. *Mol Nutr Food Res*, 2011; 55(8):1266-1270
- ⁷⁸ Guttman N. “We didn't have it in Ethiopia.” Attitudes and beliefs of Ethiopian immigrants to Israel regarding diabetes. *The 130th Annual Meeting of APHA*, 2002; Abstract presentation.
- ⁷⁹ Davidson IW, Lloyd RS, Whorwell PJ, Wright R. Antibodies to maize in patients with Crohn's disease, ulcerative colitis and Coeliac disease. *Clin Exp Immunol*, 1979, 35:147-148.
- ⁸⁰ Lehrer SB, Reese G, Malo J-L, *et al.* Corn Allergens: IgE antibody reactivity and cross-reactivity with rice, soy, and peanut. *Int Arch Allergy Immunol*, 1999; 118:298-299.
- ⁸¹ Paulis JW and Bietz JA. Separation of alcohol-soluble maize proteins by reversed-phase high performance liquid chromatography. *J Cereal Sci*, 1986; 4:205-216.
- ⁸² Asero R, Amato S, Alfieri B, *et al.* Rice: another potential cause of food allergy in patients sensitized to lipid transfer protein. *Int Arch Allergy Immunol*, 2007; 143:69-74.
- ⁸³ Horikoshi M, Kobayashi H, Yamazoe Y, *et al.* Purification and complete amino acid sequence of a major prolamin of rice endosperm. *J Cereal Sci*, 1991; 14(1):1-14.
- ⁸⁴ Urisu A, Yamada K, Masuda S, *et al.* 16-kilodalton rice protein is one of the major allergens in rice grain extract and responsible for cross-allergenicity between cereal grains in the poaceae family. *Int Arch Allergy Immunol*, 1991; 96(3):244-252.
- ⁸⁵ Wen T-N and Luthe DS. Biochemical characterization of rice glutelin. *Plant Physiol*, 1985; 78:172-177.
- ⁸⁶ Yamada K, Urisu A, Komada H, *et al.* Involvement of rice protein 16KD in cross-allergenicity between antigens in rice, wheat, corn, Japanese millet, Italian millet. (Article in Japanese) *Arerugi*, 1991; 40(12):1485-1495.
- ⁸⁷ Racusen D and Foote M. A major soluble glycoprotein of potato tubers. *J Food Biochem*, 1980; 4(1):43-52.
- ⁸⁸ Vos-Scheperkeuter GH, De Boer W, Visser RGF, *et al.* Identification of granule-bound starch synthase in potato tubers. *Plant Physiol*, 1986; 82:411-416.
- ⁸⁹ Vos-Scheperkeuter GH, de Wit JG, Ponsteinm AS, *et al.* Immunological comparison of the starch branching enzymes from potato tubers and maize kernels. *Plant Physiol*, 1989; 90:75-84.
- ⁹⁰ Eckman J, Saini SS, Hamilton RG. Diagnostic value of food-related allergic diseases. *Allergy Asthma Clin Immunol*, 2009; 5:2.
- ⁹¹ Savage JH, Matsui EC, Skripak JM, Wood RA. The natural history of egg allergy. *J Clin Immunol*, 2007; 120(6):1413-1417.

[Top](#)

⁹² Ünsel M, Sin AZ, Ordeniz Ö, *et al.* New onset egg allergy in an adult. *J Investig Allergol Clin Immunol*, 2007; 17(1): 55-58.

⁹³ Mittag D, Vieth S, Vogel L, *et al.* Soybean allergy in patients allergic to birch pollen: Clinical investigation and molecular characterization of allergens. *J Allergy Clin Immunol*, 2004; 113:148-54.

⁹⁴ Rozenfeld P, Docena GH, Añón MC, Fossati CA. Detection and identification of a soy protein component that cross-reacts with caseins from cow's milk. *Clin Exp Immunol*, 2002; 130:49–58.

⁹⁵ Selby WS, Painter D, Collins A, *et al.* Persistent mucosal abnormalities in Celiac disease are not related to the ingestion of trace amounts of gluten. *Scand J Gastroenterol*, 1999; 34:909-914.

⁹⁶ Lerner A, Rossi TM, Park B, *et al.* Serum antibodies to cow's milk proteins in pediatric inflammatory bowel disease. Crohn's disease versus ulcerative colitis. *Acta Paediatr Scand*, 1989; 78:384-389.

⁹⁷ Sanders DS, Hurlstone DP, McAlindon ME, *et al.* Antibody negative Coeliac disease presenting in elderly people—an easily missed diagnosis. *BMJ*, 200; 330(7494):775-776.

⁹⁸ Rostami K, Kerckhaert J, Tiemessen R, *et al.* Sensitivity of antiendomysium and antigliadin antibodies in untreated Celiac disease: disappointing in clinical practice. *Am J Gastroenterol*, 1999; 94:888-894.

⁹⁹ Dickey W, Hughes DF, McMillan SA. Reliance on serum endomysial antibody testing underestimates the true prevalence of Coeliac disease by one fifth. *Scand J Gastroenterol*, 2000; 35: 181-183.

¹⁰⁰ Tursi A, Brandimarte G, Giorgetti G, *et al.* Low prevalence of antigliadin and anti-endomysium antibodies in subclinical/silent Coeliac disease. *Am J Gastroenterol*, 2001; 96:1507-1510.

¹⁰¹ Tursi A, Brandimarte G, Giorgetti G. Prevalence of anti-tissue transglutaminase antibodies in different degrees of intestinal damage in Celiac disease. *J Clin Gastroenterol*, 2003; 36:219-221.

¹⁰² Abrams JA, Diamone B, Rotterdam H, Green PHR. Seronegative Celiac disease: increased prevalence with lesser degrees of villous atrophy. *Dig Dis Sci*, 2004; 49:546-550.

¹⁰³ Breiteneder H and Ebner C. Molecular and biochemical classification of plant-derived food allergens. *J Allergy Clin Immunol*, 2000; 106(1 Pt 1):27-36.

¹⁰⁴ Dubois PC and van Heel DA. Translational mini-review series on the immunogenetics of gut disease: immunogenetics of Coeliac disease. *Clin Exp Immunol*, 2008; 153(2):162–173.

¹⁰⁵ Plenge R. Unlocking the pathogenesis of Celiac disease. *Nat Genet*, 2010; 42(4):295-302.

¹⁰⁶ Ivarsson A, Hernell O, Stenlund H, ÅkePersson L. Breast-feeding protects against Celiac disease. *Am J Clin Nutr*, 2002; 75:914–921.

[Top](#)

- ¹⁰⁷ Norris J, Barriga K, Hoffenberg EJ, *et al.* Risk of Celiac disease autoimmunity and timing of gluten introduction in the diet of infants at increased risk of the disease. *JAMA*, 2005; 293:2343–2351.
- ¹⁰⁸ Naiyer AJ, Green PHR. How important is the timing of gluten introduction for children with Celiac disease? *Nat Clin Pract Gastroenterol Hepatol*, 2005; 2(10):444-445.
- ¹⁰⁹ Betterle C and Zanchetta R. Update on autoimmune polyendocrine syndromes (APS). *Acta Biomed*, 2003; 74:9-33.
- ¹¹⁰ Verdu EF, Mauro M, Bourgeois J, Armstrong D, Clinical onset of Celiac disease after an episode of *Campylobacter jejuni* enteritis. *Can J Gastroenterol*, 2007; 21(7):453-455.
- ¹¹¹ Tursi A, Brandiman G, Giorgelli GM. High prevalence of small intestinal bacterial overgrowth in Celiac patients with persistence of gastrointestinal symptoms after gluten withdrawal. *Am J Gastroenterol*, 2003; 98(4):720-722.
- ¹¹² Bonds R, Midoro-Horiuti T, Goldblum R. A structural basis for food allergy: the role of cross-reactivity. *Curr Opinion Allergy Immunol*, 2008; 8:82-86.
- ¹¹³ Green P, Alaedini A, Sander HW, *et al.* Mechanisms underlying Celiac disease and its neurologic manifestations. *Cell Mol Life Sci*, 2005; 62:791–799.
- ¹¹⁴ Jones R and Sleet S. Easily missed? *BMJ*, 2009; 338:a3058
- ¹¹⁵ Jones S, D'Souza C, Haboubi N. Patterns of clinical presentation of adult Coeliac disease in a rural setting. *Nutri J*, 2006; 5:24.
- ¹¹⁶ Feighery C. Clinical review: fortnightly review Coeliac disease. *BMJ*, 1999; 319:236-239.
- ¹¹⁷ Fasano A. Clinical presentation of Celiac disease in the pediatric population. *Gastroenterology*, 2005; 128:S68–S73.
- ¹¹⁸ Hill ID, Direks M.H Liptak GS, *et al.* Guideline for the diagnosis and treatment of Celiac disease in children: recommendations of the North American society for pediatric gastroenterology, hepatology and nutrition. *J Pediatr Gastroenterol Nutr*, 2005; 40(1):1-19.
- ¹¹⁹ Scott H, Fausa O, Ek J, Brandtzaeg P. Immune response patterns in Coeliac disease. Serum antibodies to dietary antigens measured by an enzyme linked immunosorbent assay (ELISA). *Clin Exp Immunol*, 1984; 57(1):25-32.

[Top](#)